

Organic Soil Amendment and Tillage Affect Soil Quality and Plant Performance in Simulated Residential Landscapes

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Abstract. The urban soil environment is usually not conducive to healthy root growth and function, leading to problems with plant establishment, growth, and aesthetic quality. The objective of this study was to determine if the addition of compost with or without the application of shallow tillage or aeration will improve soil physical and chemical properties and plant growth compared with an unamended control in simulated new residential landscapes. Twenty-four mixed landscape plots were established in a randomized complete block design to simulate new residential landscapes. Each plot was constructed using 10 cm of subsoil fill material over a compacted field soil and planted with *Stenotaphrum secundatum* and mixed ornamental plant species. Composted dairy manure solids were applied as an organic soil amendment at a depth of 5 cm (≈ 256 Mg-ha⁻¹) in combination with two mechanical soil treatments (tillage to 15 cm and plug aeration) for a total of five soil management treatments plus an untreated control. Soil physical and chemical properties, plant growth, and quality and plant tissue nutrient concentrations were assessed periodically to determine the effect of soil treatment on soil and plant quality. Applications of compost to soils significantly reduced soil bulk density and pH and increased soil organic matter, electrical conductivity, and Mehlich-1 phosphorus and potassium concentrations. All ornamental plant species, with the exception of *Raphiolepis indica* (L.) Lindl. ex Ker Gawl., exhibited more growth when grown in soils amended with composted dairy manure solids. In most instances, plant tissue nitrogen and phosphorus concentrations were higher for plants grown in soils receiving compost. Results of our study suggested that the addition of composted dairy manure solids to soils can improve soil properties and enhance plant growth in residential landscapes when sandy fill soils are used. In contrast, shallow tillage and aeration had little effect on soil properties or plant growth.

In recent years, many areas of the United States have experienced rapid population growth and urbanization. When land is urbanized, natural ecosystems are replaced by roads, homes, and commercial structures (Wickham

et al., 2002), often resulting in significant disturbance to soils. Studies have shown that urban soils often lack natural soil horizons (Jim, 1998), are significantly compacted (Gregory et al., 2006; Jim, 1998), can have alkaline pH (Jim, 1998; Law et al., 2004), and contain low amounts of soil organic matter (OM) and nutrients (particularly nitrogen and phosphorus) (Jim, 1998). As a result, the urban soil environment is usually not conducive to healthy root growth and function, which could lead to problems with plant establishment and growth (Cogger, 2005; Smith et al., 2001; Watson and Kelsey, 2005; Zhang et al., 2005). The management of urban soils often requires a different approach than is applied to natural or agricultural soils (Kaye et al., 2006). Some management practices that are commonly used in agricultural systems (e.g., organic amendments, shallow tillage) have the potential to improve the quality of urban soils.

The addition of organic amendments such as compost or manure to soils has been shown

to improve soil function by increasing water-holding capacity, porosity, and surface area (Cogger, 2005; Zhang, 1994). Organic amendments can help to stabilize soil structure (Thomas et al., 1996) and decrease soil bulk density (D_b) (Curtis and Claassen, 2009), thereby providing an environment that will allow for the growth of healthy root systems. The use of organic amendments during establishment of *Poa pratensis* L. has been shown to enhance turf growth and quality (Landschoot and McNitt, 1994; Linde and Hepner, 2005). Organic amendments also supply nutrients to growing plants and increase the concentrations of plant-available nutrients in soils. For example, Ingelmo et al. (1998) reported an increase in soil mineral nitrogen (N) concentration in field soils amended with sewage sludge or municipal solids waste compost (467 and 251 mg·kg⁻¹, respectively) compared with an unamended soil (79 mg·kg⁻¹). Johnson et al. (2006) reported an increase in soil phosphorus (P) and potassium (K) concentrations as compost application rate increased when dairy cattle manure compost was applied at rates of 0, 33, 66, and 99 m³·ha⁻¹. Similarly, Landschoot and McNitt (1994) showed that incorporation of various composts (e.g., biosolids, brewery byproducts, chicken manure, yard waste, horse/cow manure, papermill byproducts, mushroom substrate) into soils resulted in an increase in available P from 50.4 kg·ha⁻¹ initially to 93 to 1708 kg·ha⁻¹ depending on compost type.

The addition of organic amendments can also affect other soil chemical properties, such as pH and electrical conductivity (EC). Soil EC tends to be higher in compost-amended soils, but the effect of compost on soil pH (raising or lowering pH) is dependent on the chemical properties of the soil and the compost material itself. Calcium carbonates found in manures have been shown to increase pH when applied to slightly acid to near neutral soils (Eghball, 1999; Weindorf et al., 2006). Ginting et al. (2003) reported that the pH and EC of soils amended with beef cattle manure or composted feedlot manure (mean pH = 6.5, EC = 0.49 dS·m⁻¹) were consistently higher than soils fertilized with inorganic fertilizers or unamended soils (mean pH = 6.2, EC = 0.34 dS·m⁻¹). However, other studies have shown no change in soil pH (Foshee et al., 1999) or a decrease in soil pH (Himelick and Watson, 1990; Scharenbroch, 2009; Wright et al., 2007) after addition of organic amendments (including composts and mulches) compared with unamended soils. Wright et al. (2007) also noted that compost added to the soil provided a buffering effect to pH increases that were observed in the control as a result of irrigation water source.

Tillage can be used to improve the physical properties of compacted soils. In compacted soils, tillage breaks up massive structure, thereby increasing soil pore space and allowing water to infiltrate and roots to penetrate through the soil profile (Lipiec and Stepniowski, 1995). da Silva et al. (1997) reported that relative D_b , which is the ratio of the D_b of a soil to the D_b under standard compaction treatment (i.e.,

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samples compacted with 200 kPa of pressure), was lower in soils receiving conventional tillage (0.79 g·cm⁻³) when compared with no-till soils (0.87 g·cm⁻³). Studies have also shown that surface compaction can be alleviated by spike and core aeration or rototilling (Jim, 1993; Kozłowski, 1999; Unger and Kaspar, 1994), although the benefits of these treatments may be short-lived (Bishop and Grimes, 1978; Murphy et al., 1993). Deep tillage (to ≈0.2 to 0.4 m) promotes root growth into subsoil horizons that have more soil structural development and higher soil water-holding capacity than surface soils (Adeoye and Mohamedsaleem, 1990; Akinci et al., 2004; Busscher et al., 2006). However, Bulmer et al. (2007) reported that the benefits of deep tillage alone were temporary. When tillage to a depth of 0.75 m was applied to a field plot for growth of *Pinus contorta* Dougl., soil mechanical resistance was in excess of 2500 kPa after 1 year. However, the sandy loam soil had significantly lower soil mechanical resistance after 1 year when composted wood waste was applied in addition to tillage (Bulmer et al., 2007). Additionally, Sommerfeldt and Chang (1985) noted that there were no differences in soil D_b when compost (cattle feedlot manure) was incorporated into clay loam field soil by plow, rototiller, or cultivator.

Although research has shown that soil management practices such as organic amendment additions or tillage can improve the physical and chemical properties of soil, much of the research has been conducted in agricultural systems (Martens and Frankenberger, 1992; Roy et al., 2010) or with trees (Rivenshield and Bassuk, 2007; Scharenbroch, 2009). It is not known if these management practices can significantly improve soil conditions in urban settings, specifically new residential areas, where disturbance of the soil may contribute to environmental degradation and result in poor landscape plant growth. The objective of this study was to determine if the addition of compost with or without the application of shallow tillage or aeration will improve soil physical and chemical properties and plant growth compared with an unamended control in simulated new residential landscapes.

Materials and Methods

Experimental design. Twenty-four mixed landscape plots (3.05 m × 3.66 m) were established in a randomized complete block design at the University of Florida–Institute of Food and Agricultural Sciences (IFAS) Gulf Coast Research and Education Center in Wimauma, FL, to simulate new residential landscapes. All vegetation was removed from the site before plot construction. The entire research area was prepared at a 2% grade (as is typically required by construction codes) and compacted (D_b range: 1.7 to 1.9 g·cm⁻³) using a small plate compactor (Wacker Neuson, Munich, Germany). Individual landscape plots were constructed inside water-sealed treated wooden boxes. Within each plot, the compacted field soil (Zolfo fine sand; sandy, siliceous, hyperthermic Oxyaquic Alorthods)

(USDA-NRCS, 2004) was then buried under 1.13 m³ of uncompacted soil fill material. Three fill materials were mixed in equal parts to simulate a “top soil” material that would be applied during residential construction. The three fill soil material sources included: a subsoil fill containing construction material and other debris; a clean topsoil material (St. Johns fine sand; sandy, siliceous, hyperthermic Typic Alaquod) obtained from depth of 30 to 60 cm (Hills Dirt Pit, LLC., Riverview, FL), and a clean subsoil fill (St. Johns fine sand) fill obtained from a depth of 122 to 213 cm (Hills Dirt Pit, LLC.).

Composted dairy manure solids (compost; Agrigy, Palm Harbor, FL) were applied as an organic soil amendment at a rate of 508 m³·ha⁻¹ (5-cm depth, equaling ≈256 Mg·ha⁻¹) in combination with two mechanical soil treatments (shallow tillage and aeration) for a total of five soil management treatments: 1) tillage only; 2) compost only; 3) compost + tillage; 4) aeration only; and 5) compost + aeration. Chemical analysis of the compost indicated that the material had a pH of 6.59, an EC of 1.02 dS·m⁻¹, and a total carbon to N (C:N) ratio of 13.6. Plots that received compost applications received total N and EPA 3050 digestible P at a rate of ≈3277 and 1385 kg·ha⁻¹, respectively. In plots receiving the tillage treatment, soil (or soil + compost) was turned to a depth of 10 to 15 cm using a counterrotating tines tiller (Sears Brands, LLC, Hoffman Estates, IL). In plots receiving the aeration treatment, soil aeration plugs were mechanically removed using a core aerator (Billy Goat Industries, Inc., Lee’s Summit, MO). An untreated control plot (no tillage or organic amendment) was included as the sixth soil treatment.

Once soil treatments were applied, each plot was split across the contour and 5.58 m² of the plot was planted with *Stenotaphrum secundatum* (Walter) Kuntze turfgrass; the remaining 5.58 m² was planted with ornamental plants. Ornamental plants species, selected to represent species commonly installed in Florida urban landscapes, included: *Galphimia glauca* Cav., *Rhaphiolepis indica* (L.) Lindl. ex Ker Gawl., *Ilex cornuta* ‘Burfordii’ Lindl. & Paxton, and *Liriope muscari* (Decne.) L. H. Bailey. In all landscape plots, turfgrass was fertilized at a total N rate of 220 kg·ha⁻¹ based on current University of Florida–IFAS recommendations for South Florida (moderate maintenance) (Sartain, 2007): complete turf fertilizer (26N–0.9P–9.1K; Lesco Professional Turf Fertilizer, Sebring, FL) at an N rate of 48.8 kg·ha⁻¹ per application in February and October, polymer-coated urea (42N–0.0P–0.0K; Harrell’s Professional Fertilizer Solutions, Lakeland, FL) as a slow-release N source at an N rate of 48.8 kg·ha⁻¹ per application in May and August, urea (46N–0.0P–0.0K; Potash Corp., Northbrook, IL) as a soluble N source at an N rate of 24.4 kg·ha⁻¹ in April, and 6.34 L·ha⁻¹ of ferrous sulfate (Sunniland Corporation, Sanford, FL) in July. Ornamental plants were fertilized every 3 months with urea (40N–0.0P–0.0K) at an N rate of 24.4 kg·ha⁻¹ per application based on University of Florida–IFAS recommendations for established woody

ornamentals grown in the landscape (Knox et al., 2002). The nutrient content of added compost was not considered when fertilizing turfgrass or ornamentals because compost was applied based on horticultural recommendations to improve soil conditions (Urban, 2008) rather than as a nutrient source.

The entire research plot area was equipped with a spray irrigation system, which allowed for individual landscape plots to be irrigated, as needed, based on University of Florida–IFAS recommendations (Zazueta et al., 2005). Plots were watered daily for 30 d after planting to allow for establishment of turf and ornamental plant material. Irrigation frequency was then reduced to 2 d per week based on typical watering restrictions for landscape irrigation that would be mandated in times of drought (South Florida Water Management District, 2010; St. Johns River Water Management District, 2010). Irrigation was applied for 51 min (irrigation controller run time for two irrigation events per week at a 0.13 cm·h⁻¹ application rate, assuming system efficiency of 80% and considering effective rainfall) per plot on Mondays and Thursdays starting at 0300 HR and ending at ≈0900 HR.

Soil physical and chemical properties. Soil physical and chemical properties were measured before tillage, aeration, and compost treatments were applied and then repeated every 3 months [0, 13, 27, 40, and 52 weeks after treatment (WAT)] for a period of 1 year. Soil D_b was measured on a single soil core collected from the turf and ornamental beds (two samples per plot) at 0 to 10 cm using the core method (Blake and Hartge, 1986). Ten to 15 soil cores were collected in a random pattern from the turf areas and ornamentals beds in each landscape plot using a soil probe at a depth of 0 to 10 cm and 10 to 20 cm and mixed together to form two composite samples per plot (one per depth). Composite soil samples were then air-dried at room temperature (25 ± 2 °C) and sieved to pass a 2-mm screen. Soil pH (1:2 soil to deionized water ratio), EC (1:2 soil to deionized water ratio), and OM (loss on ignition) were determined by standard methods of the University of Florida–IFAS Extension Soil Testing Laboratory (Mylavarapu, 2009). Soil moisture content at field capacity was determined by the method described in Tan (1996) and particle size was determined by the hydrometer method (Bouyoucos, 1962). Composite soil samples were extracted using the Mehlich-1 solution (1:4 ratio of soil to 0.0125 M H₂SO₄ + 0.05 M HCl) (Mylavarapu, 2009) and analyzed for P and K by inductively coupled plasma atomic emission spectroscopy (ICP-AES).

Plant growth, root growth, and tissue analysis. Plant growth measurements and tissue nutrient content were determined at 13 and 40 WAT to evaluate the effect of soil tillage or compost amendment on the establishment and growth of ornamental plants and turfgrass. Growth index (GI) was used as a quantitative indicator of ornamental plant growth rate and to compare the size of the plants grown in the different soil treatments. GI for each plant was calculated as: GI (m³) = H × W1 × W2; where

H is the plant height (m), W1 is the widest width (m), and W2 is the width perpendicular to the widest width (m) (Scheiber et al., 2007). Turfgrass was mowed on an as-needed basis with most mowing events occurring during the summer months. Turf clippings were collected during mowing events at 13 and 40 WAT to determine clipping dry weight based on the method outlined by Ervin and Koski (2001) with some modifications. A 0.46-m wide section from the center of each plot was mowed to a height of 5.7 cm. The clippings were collected from a bag attached to the mower after every plot and then dried to a constant mass at 105 °C and weighed. Ornamental and turf tissue samples were collected by randomly sampling ≈40 to 50 leaves or blades of grass from each plot at 13 and 40 WAT. Plant tissue samples were dried at 105 °C and digested using the standard method of the University of Florida–IFAS Extension Soil Testing Laboratory (Mylavarapu, 2009) and analyzed for total Kjeldahl N (TKN), and total P and K by ICP-AES.

Shrub root cross-sectional area was used to compare the influence of soil treatments on root growth of shrub species (i.e., *G. glauca*, *R. indica*, and *I. cornuta*) based on the methods outlined by Gilman et al. (2010). Shrubs from all landscape plots were dug from the ground and soil was removed from the root balls using a high-pressure water spray. A caliper was used to measure the diameter of the 15 largest roots growing at a soil depth of 0 to 10 cm that measured greater than 1 cm. The cross-sectional area of each root was then calculated from root diameter as: CSA (cm²) = (1/2 × root diameter)² × π.

Data analysis. The experiment was designed as a randomized complete block split-plot design with four blocks and six soil treatments (main plot) in each block. Half of each plot was planted with ornamental plants and the other half was planted with turfgrass (subplot) as described previously. The soil treatments were assigned randomly within each block. Soil properties were analyzed using the PROC MIXED procedure in SAS with soil treatment as a fixed effect and block as a random effect (SAS Institute, 2003). Plant cover was added as a fixed effect for analysis of D_b samples, because separate samples were collected from turf and ornamental beds. Plant GI was analyzed separately for each shrub species using the PROC MIXED procedure in SAS with soil treatment as a fixed effect and block as a random effect (SAS Institute, 2003); initial GI (measured at 0 WAT) was included in the model as a covariate to account for variation in initial plant size. Soil and plant GI data were analyzed separately for each sample collection date when significant treatment × date interaction or date effects were noted. Root cross-sectional area was analyzed using the PROC GLM procedure in SAS (SAS Institute, 2003). All data were checked for normality by examining histogram and normality plots of the conditional residuals (generated by the command plots = residual panel). All pairwise comparisons were completed using the Tukey

honestly significant difference test with a significance level of $\alpha = 0.05$.

Results and Discussion

Soil physical and chemical properties. Initial soil samples were divided between the topsoil fill (0 to 10 cm) and the native field soil (10 to 20 cm). Soil particle size analysis indicated that the texture classification of the topsoil fill was loamy sand and the native field soil was sand. The pH was 7.5 and 6.5 and the EC was 0.30 and 0.49 dS·m⁻¹ for the topsoil fill and the field soil, respectively. Initial Mehlich-1 nutrient content analysis indicated that the field soil had lower nutrient concentrations, with the exception of P, than the topsoil fill. Mehlich-1 P and K concentrations were 145 and 20.2 mg·kg⁻¹ in the topsoil fill and 77.6 and 9.7 mg·kg⁻¹ in the native field soil. Based on soil test results, only applications of N and K would be recommended for these soils (Kidder et al., 1998).

There were no significant changes in soil particle size distribution as a result of soil treatment throughout the study; the soils were predominantly sand (mean = 88%) with very little silt (mean = 5%) or clay (mean = 6%) (data not shown). A significant soil treatment × vegetative cover interaction on soil D_b was evident; soil D_b generally followed the trend: compost-amended soils planted with ornamental plants < compost-amended soils planted with turf < unamended (control) or tilled/aerated soils planted with turf ≈ unamended (control) or tilled/aerated soils planted with ornamental plants (Table 1). Multiple researchers have reported decreased D_b as a result of incorporating compost (derived from materials including yard waste, biosolids, brewery byproducts, chicken manure, horse/cow manure, papermill byproducts, mushroom substrate) into field soils (Curtis and Claassen, 2009; Landschoot and McNitt, 1994). While we reported a decrease in soil D_b , the D_b of the unamended soils was less than the 1.8 g·cm⁻³ threshold for root restriction for a sandy soil (Hanks and Lewandowski, 2003), suggesting that D_b was unlikely to influence plant growth in our study. The significant vegetative cover × soil treatment effect on soil D_b is likely related to physiological differences in the root systems

of woody ornamentals versus turfgrass (i.e., woody versus fibrous). We hypothesize that enhanced growth of woody roots in the compost-amended soils created more macropores, resulting in decreased bulk density. Although turf root growth was probably also enhanced by application of compost, the fibrous nature of the root systems made turf roots less effective at increasing soil porosity and, thereby, decreasing bulk density.

Addition of compost (incorporated or as a top-dress) significantly increased the soil field moisture capacity compared with unamended soils at all sampling dates (Table 2). We found that compost application improved soil field moisture capacity, thereby increasing the volume of plant-available water in the soil. Pandey and Shukla (2006) reported that the application of yard trimming compost to soils at a commercial vegetable farm at a rate of 100 Mg·ha⁻¹ increased the soil moisture content compared with soils to which no compost was applied. Similarly, Curtis and Claassen (2009) found that plant-available water was increased after the addition of composted yard waste to coarse-textured field soils at a rate of 540 Mg·ha⁻¹ (on a dry mass basis; equivalent to 25% by vol.). However, the authors noted that there was no difference in plant-available water between fine-textured field soils amended with compost and unamended soils.

As expected, the application of compost increased the soil OM content compared with unamended soils through 40 WAT (Table 2). These results are consistent with many other studies that report increases in soil OM after the addition of compost materials (Curtis and Claassen, 2009; Ingelmo et al., 1998; Landschoot and McNitt, 1994; Wright et al., 2007). Manures and manure-based compost typically contain lower amounts of lignin and higher amounts of cellulose (and other more labile C forms) than composts derived from woody plant materials (Casale et al., 1995; Litvany and Ozores-Hampton, 2002), suggesting that C in these composted manures would be a readily available energy source for soil microbes. Therefore, we hypothesize that the low lignin content of the composted dairy manure solids used in our study, coupled with the warm, wet conditions that are common in Florida, allowed for rapid oxidation of C after

Table 1. Bulk density (D_b) of fill soil samples ($n = 4$) collected from 0- to 10-cm depth in simulated residential landscape plots receiving compost, shallow tillage, and/or aeration treatments averaged over five sampling dates.

Treatment	Plant cover	Bulk density (g·cm ⁻³)
Control	Ornamental plants	1.65 a ²
Tillage only	Ornamental plants	1.63 ab
Aeration only	Ornamental plants	1.66 a
Compost only	Ornamental plants	1.00 g
Compost + tillage	Ornamental plants	1.22 ef
Compost + aeration	Ornamental plants	1.09 fg
Control	<i>S. Secundatum</i>	1.52 abc
Tillage only	<i>S. Secundatum</i>	1.58 abc
Aeration only	<i>S. Secundatum</i>	1.50 abc
Compost only	<i>S. Secundatum</i>	1.46 bcd
Compost + tillage	<i>S. Secundatum</i>	1.29 de
Compost + aeration	<i>S. Secundatum</i>	1.43 cd

²Values with the same letter are not significantly different at $P < 0.05$ using Tukey's honestly significant difference test.

Table 2. Selected physical and chemical properties of fill soil samples (n = 4) collected from 0- to 10-cm depth in simulated residential landscape plots receiving compost, shallow tillage, and/or aeration treatments at five sampling dates.

Treatment	0 WAT ²	13 WAT	27 WAT	40 WAT	52 WAT
<i>Field capacity (g·kg⁻¹)</i>					
Control	105 bc ^y	123 b	123 cd	126 cd	128 c
Tillage only	101 c	120 b	119 cd	121 cd	126 c
Aeration only	101 c	123 b	107 d	111 d	118 c
Compost only	213 a	199 a	145 b	153 ab	159 b
Compost + tillage	175 a	199 a	168 a	172 a	190 a
Compost + aeration	162 ab	199 a	137 bc	139 bc	155 b
<i>Organic matter (g·kg⁻¹)</i>					
Control	16.5 b	23.1 b	15.0 ab	7.00 c	26.1 a
Tillage only	19.0 b	28.6 b	8.50 b	10.5 c	17.5 a
Aeration only	17.5 b	28.7 b	10.0 b	13.5 bc	18.4 a
Compost only	46.3 a	63.1 a	31.7 ab	32.6 ab	30.3 a
Compost + tillage	54.4 a	60.9 a	37.6 a	46.3 a	32.0 a
Compost + aeration	44.8 a	60.9 a	33.0 ab	33.1 ab	31.4 a
<i>pH</i>					
Control	7.46 a	7.63 a	7.73 a	7.66 a	7.87 ab
Tillage only	7.37 a	7.59 ab	7.76 a	7.71 a	7.98 a
Aeration only	7.48 a	7.69 a	7.86 a	7.76 a	7.93 ab
Compost only	7.32 a	7.20 c	7.11 b	7.09 b	7.39 c
Compost + tillage	7.36 a	7.36 abc	7.35 b	7.32 b	7.65 bc
Compost + aeration	7.35 a	7.22 bc	7.16 b	7.13 b	7.43 c
<i>Electrical conductivity (dS·m⁻¹)</i>					
Control	0.30 b	0.30 bc	0.13 bcd	0.40 a	0.22 a
Tillage only	0.32 b	0.29 bc	0.10 d	0.31 a	0.35 a
Aeration only	0.31 b	0.25 c	0.13 cd	0.22 a	0.41 a
Compost only	0.58 ab	0.39 a	0.18 ab	0.50 a	0.29 a
Compost + tillage	0.63 a	0.40 ab	0.16 abc	0.42 a	0.38 a
Compost + aeration	0.59 ab	0.35 a	0.20 a	0.27 a	0.41 a
<i>Phosphorus (mg·kg⁻¹)</i>					
Control	141 ab ^y	127 c	146 c	166 bc	154 bc
Tillage only	157 ab	150 bc	167 bc	180 abc	167 abc
Aeration only	130 b	135 c	172 bc	147 c	122 c
Compost only	181 a	200 a	219 a	216 ab	214 a
Compost + tillage	184 a	191 ab	219 a	217 a	193 ab
Compost + aeration	171 ab	208 a	193 ab	226 a	212 a
<i>Potassium (mg·kg⁻¹)</i>					
Control	23.3 c	27.8 b	19.8 b	23.5 a	24.0 a
Tillage only	29.3 c	35.3 b	23.8 ab	35.3 a	24.5 a
Aeration only	86.5 bc	32.5 b	21.3 b	27.5 a	27.0 a
Compost only	212 ab	64.3 a	29.8 ab	39.0 a	25.0 a
Compost + tillage	268 a	61.0 a	37.3 a	38.3 a	29.5 a
Compost + aeration	233 ab	55.8 a	39.5 a	37.8 a	29.8 a

²WAT = weeks after treatment.

³Values within the same sampling date (WAT) with the same letter are not significantly different at $P < 0.05$ using Tukey's honestly significant difference test.

the compost was applied to the soil. As a result, we reported no treatment differences on soil OM content after 40 WAT (Table 2).

The soil pH and EC were affected by the application of compost and, in some cases, tillage treatments. Soils that received compost additions had a lower soil pH (mean pH = 7.29) than unamended soils (pH = 7.70); this trend persisted from 13 through 52 WAT (Table 2). The decrease in soil pH after application of compost was the result of the addition of compost with a lower pH than reported in the fill soils (compost pH = 6.59; fill soil pH = 7.50) or the production of H₂CO₃ during microbial oxidation of compost derived C (Scharenbroch, 2009). Other researchers have documented that the pH of compost can influence soil pH when compost is applied to the soil (Eghball, 1999, 2002; Scharenbroch, 2009; Weindorf et al., 2006). The decrease in pH after addition of compost to our soils may

improve plant growth and quality because of enhanced plant availability of micronutrients (e.g., iron, manganese, copper). However, the pH of the composted soils in our study remained higher than the target pH for *S. secundatum* and woody ornamentals (6.5 and 6.0, respectively) (Kidder et al., 1998).

In general, soils receiving compost exhibited significantly higher EC than soils receiving no compost additions through 27 WAT (Table 2). Johnson et al. (2006) showed that amending soils with composted dairy manure increased soil EC when the compost application rate exceeded 99 m³·ha⁻¹; the compost application rate in our experiment was 508 m³·ha⁻¹. Stamatiadis et al. (1999) also reported an increase in soil EC after application of compost as a result of the presence of salts (other than nitrates) in the compost material. The increase in soil EC in our study was a result of the compost having

a higher EC (1.02 dS·m⁻¹) than the fill soil (0.30 dS·m⁻¹). However, the application of this compost did not increase soil EC to levels (greater than 3 dS·m⁻¹) that would be detrimental to even the most salt-sensitive landscape plants (Miyamoto et al., 2004). After 27 WAT, compost-amended soils no longer had higher EC than the soils receiving no compost, which was probably a result of the added salts leaching downward through the soil profile with heavy rain events. Additionally, Mehlich-1 sodium (Na) concentrations in the compost-amended soils decreased from 66.9 mg·kg⁻¹ at 0 WAT to 23.6 mg·kg⁻¹ (not significantly different from unamended soils) by 13 WAT. These soil test Na concentrations were below values that would impact plant growth (E.A. Hanlon, personal communication).

The nutrient content of soil was also affected by the addition of compost. Concentrations of Mehlich-1 P were generally higher in composted soils (Table 2); however, all soils in our study had very high Mehlich-1 P concentrations and would require no additional P fertilizer (Kidder et al., 1998). Wright et al. (2007) reported an increase in NH₄OAc-EDTA-extractable P in soils with increasing compost application rates. Gilley and Eghball (2002) also found that soil test (Bray 1) P at 0 to 5 and 5 to 15 cm was significantly greater after 4 years of corn production when composted beef manure was applied based on crop N requirements. Warman et al. (2009) found that an application of municipal solid waste compost applied to field soil at an N rate of 400 kg·ha⁻¹ resulted in greater Mehlich-3-extractable P and K compared with an unamended field soil. Similarly, we also showed an increase in the concentration of Mehlich-1 K concentrations when compost was applied (Table 2). Mehlich-1 soil test interpretations for Florida indicated that the soil test K in the compost amended soils was high (61 to 125 mg·kg⁻¹; no fertilizer recommended) (Kidder et al., 1998) through 13 WAT as a result of high K content in the compost. As a result, Mehlich-1 K concentrations were sufficient for growth of turf and ornamental plants throughout much of the study (Kidder et al., 1998). However, Mehlich-1 K concentrations in compost-amended soils had declined by 52 WAT, suggesting that K was absorbed by plant roots or leached downward into the soil profile (Table 2).

Plant growth, root growth, and tissue analysis. The response of plants to the application of compost was species-dependent. Although compost and tillage treatments had no effect on plant GI at 13 WAT (Table 3), one or more of the soil treatments that included compost increased the GI of *G. glauca*, *I. cornuta*, and *L. muscari* by 40 WAT (Table 3). The application of compost also increased root growth of *G. glauca* and *I. cornuta*. Mean total cross-sectional area of *G. glauca* roots was 578, 681, and 809 cm² in composted soils (compost only, compost + aeration, and compost + tillage, respectively) compared with 206 cm² in the unamended control soil. Similarly, total cross-sectional area of *I. cornuta* roots

was 706, 597, and 578 cm² in composted soils (compost only, compost + aeration, and compost + tillage, respectively), compared with 339 cm² in the unamended soils. In contrast, soil treatments did not affect the GI (Table 3) or root growth (data not shown) of *R. indica*. In general, soils amended with composted dairy manure solids resulted in larger plants than unamended soils. Shoot and root growth differences between species were attributed to differences in plant growth habits and nutritional needs. The dry mass of *S. secundatum* clippings was greater from compost-amended soils compared with uncomposted soils at 13 WAT (Table 4). Although there were no significant soil treatment effects at 40 WAT, our results suggest that adding compost may help turf to establish more quickly and produce greater biomass compared with unamended soils (Table 4).

Several researchers have reported an increase in shoot and root growth of ornamental

landscape plants or turf after the application of composts. For example, a study by Rivenshield (2003) found that additions of food waste compost to compacted urban soil increased plant vigor and growth of *Acer saccharum* Marshall and *Acer saccharinum* L. trees. Curtis and Claassen (2009) reported an increase in biomass of ornamental grasses [10.8 times greater for *Elymus multisetus* (J. G. Sm.) Burt Davy and 1.6 times greater for *Nassella pulchra* (Hitc.) Barkworth] compared with control or tillage alone treatments when yard waste compost was applied to disturbed soils formed from lahar or sandstone parent material, respectively. Similarly, Caravaca et al. (2003) reported an increase in shoot biomass of 120% and 360% for *Pistacia lentiscus* L. and *Retama sphaerocarpa* (L.) Boiss. shrubs, respectively, planted into a degraded silt-loam soil amended with composted urban residue. Scharenbroch (2009) also reported an increase in shoot and root growth

across plant species and organic amendment types when organic amendments were added to soil.

There are several possible explanations for the improved growth of ornamentals and turf reported in our study. For example, the increase in field capacity (and corresponding increase in plant-available water) may have impacted root and shoot growth. However, because plant water stress was not measured in this study, we cannot definitively say whether an increase in water-holding capacity of the soil was responsible for the increase in plant growth. Alternatively, it is possible that plant growth was enhanced as a result of increased soil fertility attributable to the addition of significant amounts of plant nutrients in the compost (particularly N that was mineralized as a result of soil microbial activity stimulated by compost application). This theory is supported by our results showing an increase in tissue TKN when plants were grown

Table 3. Growth index (m³) and plant tissue nutrient content (g·kg⁻¹) for landscape ornamentals (n = 8) grown in sandy fill soils receiving compost, shallow tillage, and/or aeration treatments in simulated residential landscape plots at 13 and 40 weeks after treatment.

Treatment	Growth index (m ³)		Total Kjeldahl nitrogen (g·kg ⁻¹)		Total phosphorus (g·kg ⁻¹)	
	13 WAT ²	40 WAT	13 WAT	40 WAT	13 WAT	40 WAT
<i>G. glauca</i>						
Control	0.25 a ³	0.62 b	13.5 b	15.5 b	15.6 c	29.0 a
Tillage only	0.25 a	0.50 b	12.5 b	15.4 b	16.3 bc	27.4 a
Aeration only	0.27 a	1.42 a	13.1 b	14.6 b	17.5 bc	28.2 a
Compost only	0.27 a	1.69 a	23.3 a	31.6 a	31.3 a	28.4 a
Compost + tillage	0.31 a	0.53 b	24.1 a	22.0 ab	23.5 abc	25.5 a
Compost + aeration	0.34 a	1.38 a	20.5 a	21.5 ab	28.2 ab	26.2 a
<i>I. cornuta</i>						
Control	0.13 a	0.21 cd	16.1 bc	12.9 b	12.1 a	12.9 bc
Tillage only	0.15 a	0.21 cd	15.2 c	11.8 b	10.2 a	11.3 c
Aeration only	0.14 a	0.19 d	16.0 bc	13.0 b	11.0 a	12.4 c
Compost only	0.12 a	0.39 a	17.9 ab	18.9 a	11.8 a	16.0 a
Compost + tillage	0.13 a	0.28 bc	17.1 abc	17.1 a	10.1 a	15.5 ab
Compost + aeration	0.17 a	0.34 ab	18.9 a	16.8 a	13.1 a	16.6 a
<i>L. muscari</i>						
Control	0.02 a	0.04 b	23.9 bc	20.0 c	55.8 ab	48.1 a
Tillage only	0.02 a	0.04 b	23.3 c	19.1 c	50.6 abc	45.6 ab
Aeration only	0.02 a	0.04 b	23.4 c	20.5 bc	59.1 a	50.7 a
Compost only	0.08 a	0.08 a	26.0 ab	26.6 a	42.0 c	35.7 bc
Compost + tillage	0.08 a	0.06 ab	27.3 a	24.5 ab	42.8 bc	34.6 c
Compost + aeration	0.03 a	0.07 a	26.4 a	24.6 ab	46.9 abc	29.0 c
<i>R. indica</i>						
Control	0.08 a	0.11 a	16.0 b	15.3 b	20.7 a	25.5 ab
Tillage only	0.08 a	0.09 a	15.3 b	13.9 b	23.3 a	26.4 ab
Aeration only	0.09 a	0.11 a	15.9 b	14.1 b	19.4 a	24.4 b
Compost only	0.07 a	0.14 a	18.8 a	19.0 a	22.5 a	31.2 ab
Compost + tillage	0.06 a	0.12 a	19.0 a	19.2 a	21.0 a	25.3 ab
Compost + aeration	0.08 a	0.11 a	19.1 a	21.0 a	25.0 a	34.8 a

²WAT = weeks after treatment.

³Values within the same sampling date (WAT) with the same letter are not significantly different at $P < 0.05$ using Tukey's honestly significant difference test.

Table 4. Dry mass and nutrient concentrations (n = 4) of *Stenotaphrum secundatum* clippings collected from turf grown in sandy soils receiving compost, shallow tillage, and/or aeration treatments in simulated landscape plots at 13 and 40 WAT.

Treatment	Dry mass (g)		Total Kjeldahl nitrogen (g·kg ⁻¹)		Total phosphorus (g·kg ⁻¹)	
	13 WAT	40 WAT	13 WAT	40 WAT	13 WAT	40 WAT
Control	13.5 b	5.36 a	18.1 b ³	17.3 b	45.1 b	55.0 ab
Tillage only	14.5 b	3.97 a	18.8 b	17.2 b	44.4 b	57.4 a
Aeration only	19.4 b	5.37 a	20.3 b	16.8 b	47.5 b	58.3 a
Compost Only	64.9 a	26.6 a	28.1 a	27.2 a	61.9 a	44.7 c
Compost + tillage	86.6 a	19.6 a	28.7 a	26.1 a	68.1 a	39.1 bc
Compost + aeration	70.1 a	31.8 a	26.2 a	28.1 a	66.6 a	40.8 bc

²WAT = weeks after treatment.

³Values within the same sampling date (WAT) with the same letter are not significantly different at $P < 0.05$ using Tukey's honestly significant difference test.

in soils receiving compost (+ inorganic N) compared with those grown in unamended soils (inorganic N only) (Tables 3 and 4). The concentration of TKN in the tissue of ornamentals and turf grown in soils receiving composted was higher than for plants grown in uncomposted soils that received only inorganic sources of N (Tables 3 and 4). Based on published concentrations of adequate tissue N for woody ornamentals (20 to 25 mg·kg⁻¹) (Yeager, 2010) and *S. secundatum* (20 to 30 mg·kg⁻¹) (Sartain, 2008), our results suggest a plant response as a result of the mineralization of compost N (perhaps resulting from enhanced soil microbial activity).

Although there was a more pronounced effect of compost on tissue P content of *G. glauca* (13 WAT) and *I. cornuta* (40 WAT) than the other ornamental plant species (Tables 3 and 4), the overall trends indicate that tissue P concentrations were sufficient for plants grown with or without compost [1.5 to 5.0 mg·kg⁻¹ for woody ornamentals and *S. secundatum* (Sartain, 2008; Yeager, 2010)]. Soil treatments had little consistent effect on levels of tissue total K at any time for any shrub species, but tissue levels were above reported sufficiency ranges for all treatments (data not shown). Therefore, a response to P and K added in the compost treatments was unlikely.

Conclusions

Based on results from our study, we suggest that composted dairy manure solids can improve soil physical and chemical properties in residential landscapes when sandy fill soils are used. Application of composted dairy manure solids can also enhance the establishment and improve the growth of selected ornamental landscape plants. However, topdressing with composted dairy manure solids enhanced plant growth and quality as much as incorporation of compost to a depth of 20 cm by tillage. In contrast, shallow tillage and aeration had little effect on the physical properties (e.g., bulk density, field capacity) of sandy fill soils. Our results may have been different if finer-textured soils had been evaluated, in which the threshold for D_b above which root growth would be compromised is lower. Similarly, there were no significant effects of plug soil aeration on plant establishment or growth, suggesting that the lack of effects from soil physical disturbances (tillage or aeration) was the result of the coarse-textured soils that allowed for adequate root growth at the recorded bulk density levels. Although the results of this study can only show the benefits of compost additions during the first year after planting, the increased growth and the subsequent health of plants measured in this experiment resulting from applications of compost may prevent future plant failure. Future research should determine if improved plant growth in compost-amended soils was a result of additions of N in the compost, enhanced mineralization of compost N resulting from increased soil microbial activity, or improved soil physical properties (specifically water-holding capacity). Future

research should also evaluate the long-term effects of compost addition after the plant establishment period.

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